



## Synthesis of 1-*O*-Alkyl- and 1-*O*-Acyl-*myo*-inositol 3, 4, 5-trisphosphates as Novel Analogues of Phosphatidyl-*myo*-inositol 3, 4, 5-trisphosphate

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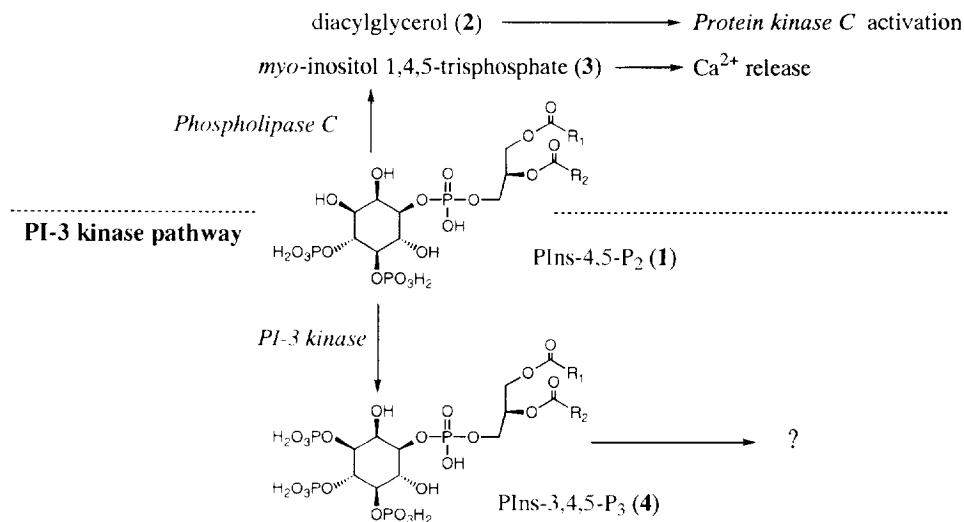
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**Abstract:** Homochiral 1-*O*-alkyl and 1-*O*-acyl derivatives of *myo*-inositol 3,4,5-trisphosphate were designed and synthesized as novel analogues of phosphatidyl-*myo*-inositol 3,4,5-trisphosphate. 1-*O*-Octadecyl and 1-*O*-stearoyl analogues inhibited the synthesis of phosphatidyl-*myo*-inositol 3,4-bisphosphate from phosphatidyl-*myo*-inositol 3,4,5-trisphosphate catalyzed by PI(3,4,5)-P<sub>3</sub> 5-phosphatase. These compounds may act as competitors in the enzyme reaction, suggesting that they may be useful in studies of the role of phosphatidyl-*myo*-inositol 3,4,5-trisphosphate as a candidate second messenger in cellular signal transduction.

Polyphosphoinositides are important in cellular signal transduction. <sup>1</sup> Phospholipase C, which is activated upon stimulation with many growth factors, hydrolyzes phosphatidyl-*myo*-inositol 4,5-bisphosphate (**1**) to produce diacylglycerol (**2**), which activates protein kinase C, and *myo*-inositol 1,4,5-trisphosphate (**3**), which releases Ca<sup>2+</sup> ions from intracellular storage sites. Recently, it has been shown that phosphatidyl-*myo*-inositol (PI) 3 kinase catalyzes phosphorylation of **1** to phosphatidyl-*myo*-inositol 3,4,5-trisphosphate (**4**)<sup>2</sup>, which is resistant to known phospholipase Cs. No enzyme that hydrolyzes **4** has been found yet, suggesting that **4** may be metabolized through a different pathway from the classical phosphatidyl-*myo*-inositol turnover. Stephenes et al. reported an enzyme that dephosphorylates phosphatidyl-*myo*-inositol 3,4,5-trisphosphate to produce phosphatidyl-*myo*-inositol 3,4-bisphosphate<sup>3</sup>, but no other enzyme that metabolizes this compound has been found so far. Although several reports on the syntheses<sup>4</sup> and roles<sup>5</sup> of **4** have appeared, the role of **4** is not yet clear. In order to obtain some informations on the role and function of **4**, we designed and synthesized the novel analogues of **4**.

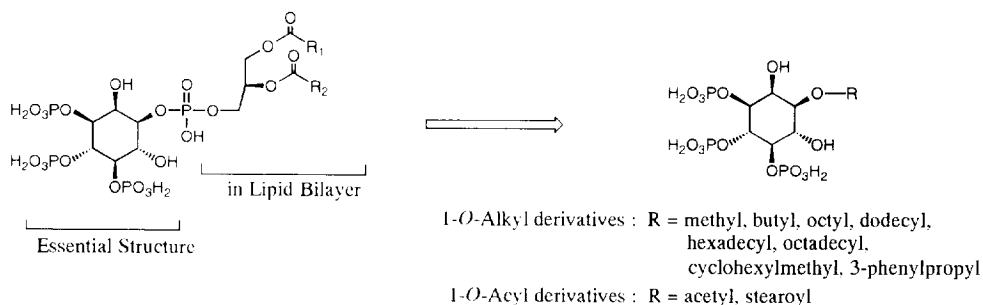
We hypothesized that the diacylglycerol moiety of **4** functions for hydrophobic interaction with the cell membrane, and could be substituted with simple lipophilic alkyl or acyl chains, and we also considered that the *myo*-inositol trisphosphate moiety facing the cytosol should play an essential role in the function of **4**. On the basis of this assumption, we synthesized a series of 1-*O*-alkyl- and 1-*O*-acyl-*myo*-inositol 3,4,5-trisphosphates as analogues of **4** (Scheme 2).

**Scheme 1**  
**Classical pathway**

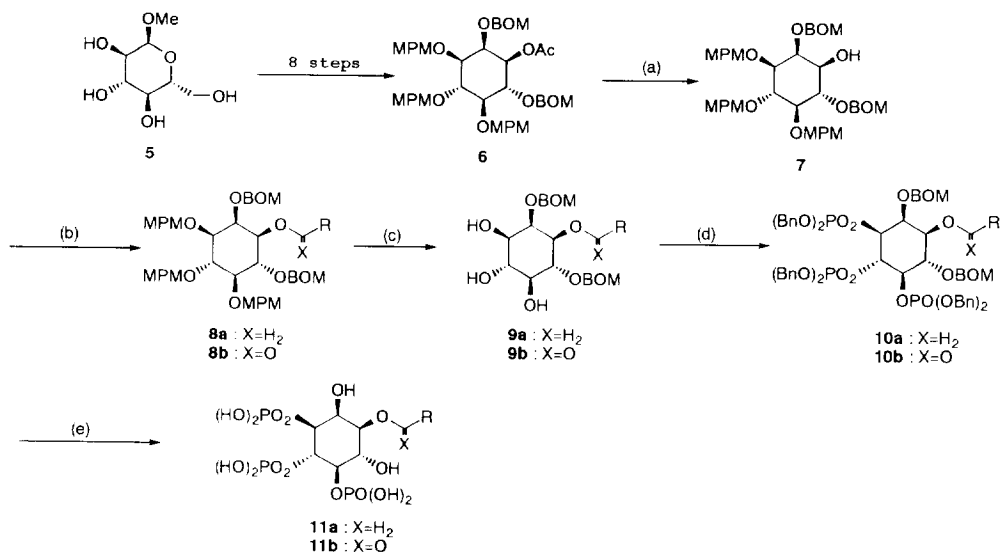


Synthesis of these 1-*O*-alkyl- and 1-*O*-acyl-*myo*-inositol 3,4,5-trisphosphates is illustrated in **Scheme 3**. The common key intermediate, 2,6-di-*O*-benzyloxymethyl-3,4,5-tri-*O*-*p*-methoxybenzyl-*myo*-inositol (7), was prepared from methyl- $\alpha$ -D-glucopyranoside according to the reported procedure.<sup>6</sup> The alcohol (7) was alkylated with the corresponding alkyl halides (methyl iodide for a 1-*O*-methyl derivative; alkyl bromide for others) in the presence of KOH in dimethylsulfoxide<sup>7</sup> or acylated with stearic anhydride in the presence of 4-(dimethylamino)pyridine in pyridine. The 1-*O*-acetyl derivative was directly obtained from 1-*O*-acetyl-2,6-

**Scheme 2**



Scheme 3

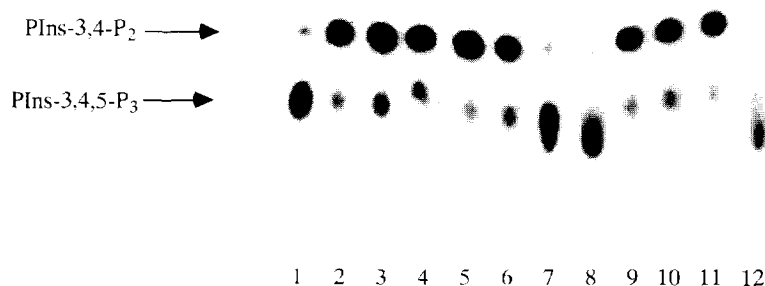


**Reagents and conditions** (a) NaOH, MeOH, reflux, 1 h (89 %); (b) for alkylation: alkyl bromide or iodide, KOH, DMSO, r.t., 18 h (25–100%); for acylation: acid anhydride, DMAP, pyridine, r.t., 36 h (51 %); (c) DDQ, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 1 h (70.5–100 %); (d) tetrazole, dibenzyl *N,N*-diethylphosphoramidite, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 30 min, then *m*CPBA, -40°C to r.t., 18 h (28–96%); (e) H<sub>2</sub>, 10% Pd-C, 50 psi, 95 % EtOH, r.t., 5.5 h, (56–100%)

di-*O*-benzyloxymethyl-3,4,5-tri-*O*-*p*-methoxybenzyl-*myo*-inositol (**6**), an intermediate for the synthesis of **7**. Removal of the *p*-methoxybenzyl groups with 2,3-dichloro-5,6-dicyano-*p*-benzoquinone (DDQ) in wet dichloromethane gave the triols (**9**). These were phosphitylated with dibenzyl *N,N*-diethylphosphoramidite and the resulting trisphosphites were oxidized with *m*-chloroperbenzoic acid to give the desired fully-protected trisphosphates (**10**). All protective groups of **10** were removed by hydrogenolysis (10% Pd/C, 50 psi) and the filtrates were neutralized with diluted ammonium hydroxide to give 1-*O*-alkyl and 1-*O*-acyl-*myo*-inositol 3,4,5-trisphosphate ammonium salts<sup>8</sup>.

To see whether or not these derivatives serve as analogues of phosphatidyl-*myo*-inositol 3,4,5-trisphosphate, we examined their effect on Plns-3,4,5-P<sub>3</sub> 5-phosphatase (**Figure 1**), recently isolated by us<sup>9</sup>. 1-*O*-Hexadecyl, 1-*O*-octadecyl and 1-*O*-stearoyl derivatives completely inhibited this enzyme activity. In contrast, the 1-*O*-dodecyl derivative showed only a weak effect and others with shorter side chains had no activity. The results suggested that the presence of a long lipophilic side chain is necessary for Plns-3, 4, 5-P<sub>3</sub> 5-phosphatase-inhibitory activity. These novel 1-*O*-alkyl and 1-*O*-acyl-*myo*-inositol 3,4,5-trisphosphates may be applicable for the preparation of biochemically useful tools, such as fluorescent probes or photoaffinity labeling reagents, for further studies of the role of phosphatidyl-*myo*-inositol 3,4,5-trisphosphate in cellular signal transduction.

Figure 1



The inhibitory activity of 1-*O*-alkyl and 1-*O*-acyl derivatives on Plns-3,4,5- $P_3$  5-phosphatase. Plns-3,4,5- $P_3$  was prepared from phosphatidyl-*myo*-inositol 4,5-bisphosphate and [ $\gamma$ - $^{32}$ P]ATP by purified PI 3-kinase. Effects of various compounds on Plns-3,4,5- $P_3$  5-phosphatase activity were assayed by using partially purified enzyme from bovine thymus (see ref. 9). The reaction mixtures (30  $\mu$ l) contained Plns-3,4,5- $P_3$  (20 nM), phosphatidylserine (200  $\mu$ g/ml), 20 mM Tris/HCl (pH 7.5), 150 mM NaCl, 1 mM  $MgCl_2$ , 0.3 % octylglucoside, partially purified enzyme, and  $PIP_3$ -analogue (0.5 mM). After incubation for 10 min at 25°C, the reaction was stopped by the addition of chloroform / methanol / HCl (200 / 100 / 1). Then, 1N HCl was added, and the mixture was separated into two phases by centrifugation. Organic phase was spotted onto a Silica Gel 60 Plate (Merck), which was developed in chloroform / methanol / acetone / acetic acid /  $H_2O$  (40 / 15 / 13 / 12 / 7). Lipids were visualized by autoradiography. Lane 1: no enzyme; 2: control; 3: methyl; 4: butyl; 5: octyl; 6: dodecyl; 7: hexadecyl; 8: octadecyl; 9: 3-phenylpropyl; 10: cyclohexylmethyl; 11: acetyl; 12: stearoyl.

## ACKNOWLEDGEMENT

We are grateful to Dr. Naoko Morisaki for measurements of negative FABMS and to Dr. Kazuo Furihata for the measurements of  $^{31}P$  NMR.

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- All synthetic intermediates were characterized by  $^1H$ -NMR, FABMS, HRMS, IR and optical rotation measurements. The final products were characterized by  $^1H$ -NMR,  $^{31}P$ -NMR, negative FABMS and optical rotation as the corresponding ammonium salts.  
Representative data for selected compound: 1-*O*-dodecyl-*myo*-inositol 3,4,5-trisphosphate,  $^1H$ -NMR: (500 MHz,  $D_2O$ )  $\delta$ : 0.76 (t, 7Hz, 3H), 1.14-1.26 (br, 18H), 1.50 (m, 2H), 3.31 (brd, 10Hz, 1H, inositol), 3.44 (brm, 1H), 3.61 (brm, 1H), 3.74 (brm, 1H, inositol), 3.89-4.00 (brm, 2H, inositol), 4.30 (brm, 1H, inositol), 4.34 (br, 1H, inositol).  $^{31}P$ -NMR: (205.5 MHz,  $D_2O$ )  $\delta$ : 0.55, 1.07, 1.34. Negative FABMS: 587 (M-H), 609 (M-2H+Na).  $[\alpha]_D^{25} = -11.8^\circ$  (c=0.30,  $H_2O$ ).
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